

## 4 ON NAMIBIA'S MARINE FISH DIVERSITY

*Gabriella Bianchi, Elizabeth Lundsør and Hashali Hamukuaya*

### *Abstract*

Conservation of marine diversity is a major challenge worldwide. Some of the difficulties reside in the paucity of baseline information on species diversity at the local level, and the lack of operational indices and reference points that may be useful for management purposes. In this study, various aspects of the diversity of Namibian demersal fish assemblages are explored, based on comprehensive datasets of fish occurrence and abundance obtained from bottom trawl surveys with the Norwegian research vessel Dr. Fridtjof Nansen. The study is meant to provide baseline information on Namibian demersal fish diversity that will hopefully represent a useful reference for future conservation efforts.

### INTRODUCTION

Conservation of biological diversity has been high on the international political agenda since the 1992 United Nations Conference on Environment and Development (UNCED). The conference generated the Convention on Biological Diversity (CBD) that entered into force in 1993. Article 7 of the CBD deals with identification and protection of biodiversity, Article 10 deals with its sustainable use while Article 12 encourages the establishment of programmes for scientific research and education ‘...in measures for the identification, conservation and sustainable use of biological diversity... and provide support for such training and education for specific needs in developing countries’.

Implementing the CBD is a global challenge. Effective biodiversity management requires detailed information on biogeographic distribution of habitats and species and clearly defined objectives and performance indicators. Ten years since the CBD entered into force, this type of information is yet not available for most ecosystems. The challenge is greatest for marine habitats, given the difficulty in studying aquatic systems and the high costs of research on these systems. Although it may be argued that marine species

are probably less vulnerable to local extinctions than terrestrial species, given that dispersal and new recruitment are easier than in terrestrial habitats, conservation of marine diversity should still be regarded as very important for the maintenance of ecosystem functions (Warwick and Clarke, 2001).

The only information available on fish diversity off Namibia is the type found in the FAO publications such as the Species Identification Sheets for Fishery Purposes (Fischer *et al.*, 1981), national field guides (Bianchi *et al.*, 1999) and species catalogues. These publications summarise published records of species occurrences and generalised charts are drawn based on these records. Table 1 presents an overview of the taxa included in the FAO national field guide for Namibia (Bianchi *et al.*, 1999), that includes the most comprehensive list of species recorded in this area. Similar information can also be found in FishBase, a global information system on fishes (Froese and Pauly, 2003). The information found in these works, however, does not have the level of detail needed for management purposes given that species distributions are indicated following broad biogeographic patterns and are not based on local habitat distribution. Also, monitoring diversity in a given area should take into account aspects of local abundance and dominance that can only be evaluated by sampling and comparing trends over time.

This study is meant to contribute to the description of biogeographic patterns of the diversity of demersal fish communities off Namibia. Furthermore, conventional and new indices of diversity will be applied to explore different aspects of fish diversity on the continental shelf and upper slope of Namibia.

## MAIN BIOGEOGRAPHIC PATTERNS

Namibia's demersal assemblages have been described in Bianchi *et al.* (1999) and Hamukuaya *et al.* (2000). Major changes in the composition of the Namibian demersal fauna take place along the depth gradient and with latitude. A major faunal boundary is found at the shelf edge, between about 300 and 350 m depth, separating shelf from slope communities. Main patterns found

Table 1. Overview of recorded fish taxa, based on Bianchi *et al.* (1999).

Taxa	Agnatha	Elasmobranchs	Bony fishes
Orders	1	9	24
Families	1	25	129
Genera	2	47	296
Species	2	83	424

on the shelf are associated with latitude and a clear boundary is found at around 27°S, i.e. around the Lüderitz upwelling cell, and another at around 21°S. Main shelf assemblages are therefore three, each characterised by a distinctive species composition and associated with different environmental conditions. The northern shelf assemblage (from the Angolan border to about 21°S) is characterised by a number of tropical/subtropical species that reach here their southernmost distribution (e.g. *Dentex macrophthalmus*, *Synagrops microlepis* and *Pterothrissus belloci*). The central shelf area, between the latitudes of about 21 and 27 °S, is dominated by *Merluccius capensis* (Cape hake) and *Sufflogobius bibarbatatus* (pelagic goby) and characterised by being the main oxygen-deficient area in the Benguela system. Near-bottom oxygen levels found here are very low, seldom > 0.5 ml l<sup>-1</sup>. Diversity here should be expected to be lower and the species found here have special adaptations to withstand these extreme environmental conditions. South of Lüderitz and to the border with South Africa, the fish community is characterised by species that reach their northernmost limit, having their main distribution off South Africa. Near-bottom oxygen levels on this part of the shelf are usually above 2.5 ml l<sup>-1</sup>. Dominating species are the hakes *Merluccius paradoxus* and *M. capensis*, *Lepidopus caudatus*, *Emmelichthys nitidus*, *Zeus capensis*, *Chelidonichthys capensis*, *Genypterus capensis*, *Thyrssites atun* and *Callorhinchus capensis*.

The upper slope assemblage (between 300 and 600 m depth) includes a few species also found on the shelf but also a number of deep-water benthic fishes. The deep-water hake *Merluccius paradoxus* is the dominating species, accompanied by several species of Macrouridae, *Helicolenus dactylopterus* and several species of deep-water sharks. Figure 1 shows the Namibian coast with the distribution of the main communities.

Demersal fish communities off Namibia have been intensively exploited since the early 1960s and, to the authors' knowledge, no comprehensive diversity studies were carried out at the early stages of exploitation. Bianchi *et al.* (2001) looked at trends in community diversity (richness, Shannon-Wiener and Simpson) and abundance/biomass comparisons (ABC plots) for the decade after Independence. The main feature that was revealed was an increase in diversity due to increased evenness during the study period in the upper slope area. An increase in richness was also observed but was attributed to improved survey protocols. Another important feature was the clear difference in richness and dominance (evenness) patterns between the shelf and the upper slope assemblages, with species richness and evenness increasing with depth.

These conventional measures, however, have some drawbacks (Clarke and Warwick, 2001a) related to their dependence on sample size/effort. Furthermore, they lack a statistical framework to test whether and to what extent a sample departs from the expectation. Finally, they do not reflect

phylogenetic diversity and therefore do not take into account to what extent species are taxonomically related. For these reasons Warwick and Clarke (1995) developed a set of indices that also take into account the taxonomic relatedness of the species present in a community. These indices have only been applied to fish communities in two works. Hall and Greenstreet (1998) compared the performance of these indices with conventional diversity indices based on species richness and evenness for a demersal fish commu-

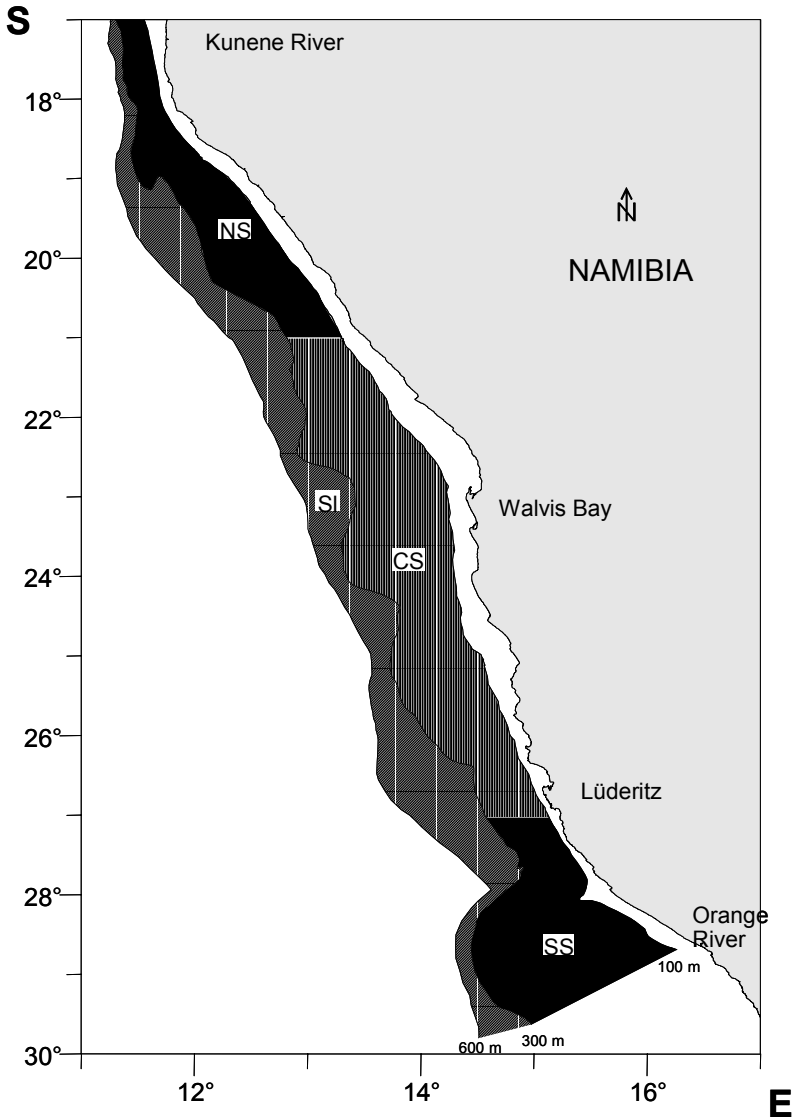


Figure 1. Namibia's shelf and upper slope area, showing the main habitats in this study. SL, Slope; NS, Northern Shelf; CS, Central Shelf; SS, Southern Shelf.

nity of the North Sea. They concluded that there was no difference in the observed trends between the conventional and the new 'taxonomic' indices but they argued that this was due to the fact that major changes had taken place in the community structure and that these were therefore well reflected in both types of indices.

Rogers *et al.* (1999) applied the new taxonomic indices on bottom-dwelling fish off NW Europe and found that greater resolution was achieved by using these indices as compared with conventional ones. They also noted that the taxonomic range, including both species redundancy and range of taxa with species fulfilling key functional roles, may be important for maintaining the ecosystem stability.

## MATERIALS AND METHODS

The data used in the current analysis were collected through bottom trawl surveys by the Norwegian research vessel Dr Fridtjof Nansen off Namibia, from 1990 to 2000 (Table 2). The stations were sampled randomly along transects perpendicular to the coast separated from each other by about 20 nautical miles and data collected from a total of 2309 stations were used for this analysis. The sampling gear was a high-opening shrimp and bottom fish trawl, with a headline of 31 m, footrope of 47 m, roller disks of 12 cm in diameter. The estimated headline height during towing was 5 m. The codend was lined with fine meshes of 20 mm. Towing lasted about 30 minutes at a speed of 3 knots.

Table 2. Overview of the surveys and number of trawl hauls used in the analysis. Project codes and station numbers, as used in the database from which the data were extracted, are also included.

Year	Start date	End date	Project code	St. Nr.	N Stations
1990	26/01	18/03	NA	2 - 242	241
1991	27/01	20/03	NA	501 - 710	210
1992	24/04	19/05	NA	1061 - 1225	165
1993	21/01	23/02	NA	1566 - 1759	194
1994	21/01	21/02	N1	1 - 152	152
1995	22/04	28/05	N1	920 - 1104	185
1996	14/01	17/02	N1	1185 - 1427	243
1997	12/01	19/02	N1	1857 - 2124	267
1998	13/01	20/02	N1	2229 - 2442	214
1999	13/01	18/02	NC	2600 - 2814	215
2000	17/01	22/02	NC	2815 - 3037	223

The catches were sampled and all species identified, on the basis of existing literature (Fischer *et al.*, 1981; Bianchi *et al.*, 1993; Smith and Heemstra, 1991). Each species was weighed and counted and catches standardised to catch per hour tow. Pelagic species belonging to the families Clupeidae, Engraulidae and Carangidae were not included in the analyses.

### Data analysis

The continental shelf and upper slope of Namibia, between 100 and 600 m, were subdivided into four areas (Figure 1 and Table 3) based on Hamukuaya *et al.* (2000). For each area, analyses were performed both on individual samples and on the average catches over the whole sampling period.

Data were exported from the NANSIS database (Strømme, 1992) and matrices were generated with species abundance by sampling station, for each of the four areas over the whole study period. Another table was generated including pooled samples representing the average species abundance in each of the four areas described above. The former were used for calculating  $\alpha$ -diversity while the latter would produce  $\gamma$ -diversity indices.  $\alpha$ -diversity is the diversity at the local scale while  $\gamma$ -diversity describes diversity on a larger scale, here the total diversity of a given area (Whittaker, 1960).

All the calculations were done using the Primer-e programme (Clarke and Warwick, 2001b) for Windows.

### Diversity indices

Since no single biodiversity index covers all aspects of biodiversity, a range is needed to describe marine communities (Jennings and Reynolds, 2000). In this work we have chosen to include the Shannon-Wiener index, species richness, Simpson's indices of dominance and evenness and newly developed indices for taxonomic diversity and distinctness. The higher taxa used for the analyses are based on the taxonomy presented by Moyle and Cech (1982), for the higher taxa (phylum to suborder) and Bianchi *et al.* (1999) for lower taxa (family to species). Fishbase (Froese and Pauly, 2003) was used for those species not included in Bianchi *et al.* (1999).

Table 3. Summary of spatial distribution of assemblages by latitude (Ang, border with Angola; SA, border with South Africa).

Area	Depth (m)	Latitude	N Stations
Upper slope	301-600	Ang-SA	1010
Northern shelf	100-300	Ang- 21°S	315
Central shelf	100-300	21- 27°S	513
Southern shelf	100-300	27°S-SA	180

The Shannon-Weiner index,  $H'$ , is a commonly used index that expresses both species richness and evenness. It gives the share of  $N$  (individuals) per  $S$  (species).

$$H' = - \sum p_i \log p_i \quad (1)$$

where  $p_i$  is the percentage of species  $i$  in relation to the total number ( $S$ ).

Species richness ( $S$ ) is often given as total number of species. Richness is dependent on sample size so that it is important to show the relationship between the total number of species versus the number of samples or individuals sampled (rarefaction curves). We have chosen to display 'area plots' to show richness (cumulative number of species sampled) as a function of number of samples collected for each of the assemblages considered. For this purpose all samples taken in a given area have been used.

Simpson's indices,  $D$  and  $E$ , express dominance and evenness of species:

$$\text{Dominance: } D = \sum p_i^2 \quad (2a)$$

$$\text{Evenness: } E = 1 - D \quad (2b)$$

None of the above indices, however, includes the higher taxonomic relationship between species. They may therefore show the same diversity values for assemblages that comprise species closely related to each other as for assemblages where the species are taxonomically more distinct. Intuitively we would consider the latter case more diverse than the former. Warwick and Clarke (1995) developed biodiversity indices that express taxonomic diversity.

Average taxonomic diversity  $\Delta$  can be considered as an extension of the Simpson diversity with the inclusion of taxonomic separation. It is described as the mean hierarchical path length between any randomly chosen individuals ( $x$ ) (Jennings and Reynolds, 2000) while reflecting dominance patterns among species.

$$\Delta = [\sum \sum_{i < j} \omega_{ij} x_i x_j] / [N(N-1)/2] \quad (3)$$

where  $N = \sum_i X_i$  i.e. all the individuals sampled and  $\omega_{ij}$  is the taxonomic distances through the classification tree, between every pair of individuals.

Taxonomic distinctness  $\Delta^*$  is the expected taxonomic distance between two randomly chosen individuals under the condition that they represent two different species (Jennings and Reynolds, 2000). The dominance effect is reduced as compared with  $\Delta$ . This is considered by far the most sensitive univariate measure of community structure by Warwick and Clarke (1995).

$$\Delta^* = [\sum \sum_{i < j} \omega_{ij} x_i x_j] / [\sum \sum_{i < j} x_i x_j] \quad (4)$$

Average taxonomic breadth  $\Delta^+$  is the taxonomic path length for presence/absence data and expresses the average taxonomic distinctness (Warwick and Clarke, 1995)

$$\Delta^+ = [\sum_{i<j} \omega_{ij}] / [S(S-1)/2] \tag{5}$$

where S is the number of species in the sample.

Clarke and Warwick (1998) have shown that these indices are largely independent of sampling effort.

**Weighing of samples**

$\omega$  is the path length between two species.  $\omega$  can be equal for each step in the taxonomic tree, (100/number of steps) or it can be based on taxon richness (Clarke and Warwick, 1999). The last approach was used in this study.

The values obtained in taxonomic breadth for samples within each community were tested against the average taxonomic breadth for all groundfish species sampled and identified during the study period, as described in Clarke and Warwick (2001b).

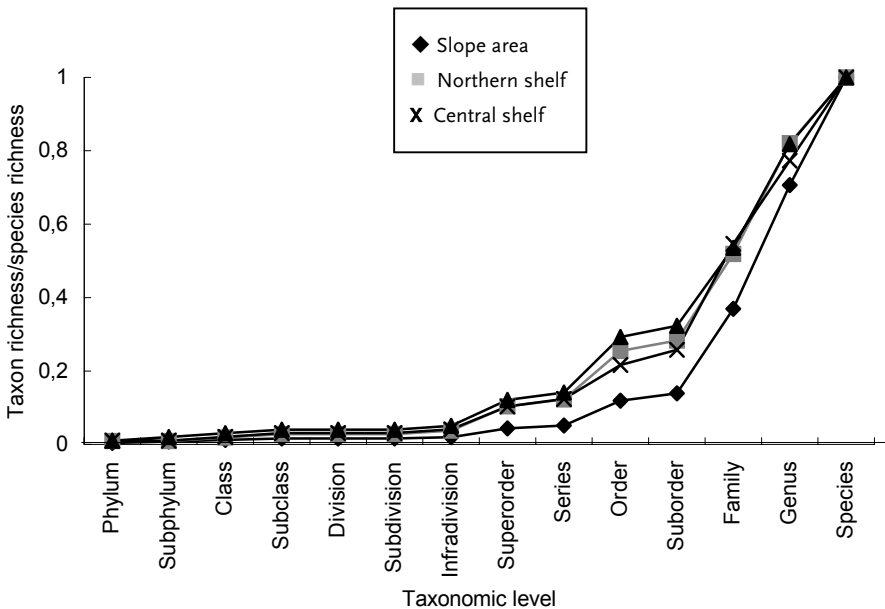


Figure 2. Profiles of ratios between richness at each taxonomic level and number of species in each community. Diamonds: Slope community; crosses: Southern shelf community; squares: Northern shelf community; triangles: Central shelf community.



## RESULTS

Appendix 1 shows the list of species and families of bottom-dwelling fishes caught in all trawl stations used in this study. Table 4 shows the number of taxa for each level of classification by main areas. The upper slope holds the highest number of species, genera and families, while at higher taxonomic levels the taxon richness values are comparable between the various communities/areas. In Figure 2 this is shown by comparing the ratio between taxon richness at all taxonomic levels used and the number of species in each area/community. The profiles show lower ratios for the slope community at higher taxonomic levels.

### $\alpha$ - diversity

A measure of local species richness is provided by the average number of species caught per trawl haul (Figure 3). There are significant differences, with the upper slope area showing the highest value, followed by the southern shelf, while the central and northern shelves have the lowest values. Figure 3 also shows trends in dominance, with the shelf communities clearly characterised by higher dominance levels (lower evenness) and the upper slope with lower dominance (higher evenness).

Figure 4 shows means and variances for the Shannon-Wiener index ( $H'$ ), average taxonomic diversity ( $\Delta$ ), distinctness ( $\Delta^*$ ) and average taxonomic breadth ( $\Delta^+$ ) for each of the communities at the  $\alpha$ - diversity level. Two of the

Table 4. Number of groups in each taxonomic level, by main area.

	Slope	Northern shelf	Central shelf	Southern shelf
Phylum	1	1	1	1
Subphylum	2	1	2	1
Class	3	2	3	2
Subclass	4	3	4	3
Division	4	3	4	3
Subdivision	4	3	4	3
Infradivision	5	4	5	4
Superorder	11	11	12	10
Series	13	13	14	12
Order	30	27	29	21
Suborder	35	30	32	25
Family	93	55	53	53
Genus	178	87	81	75
Species	252	106	99	97

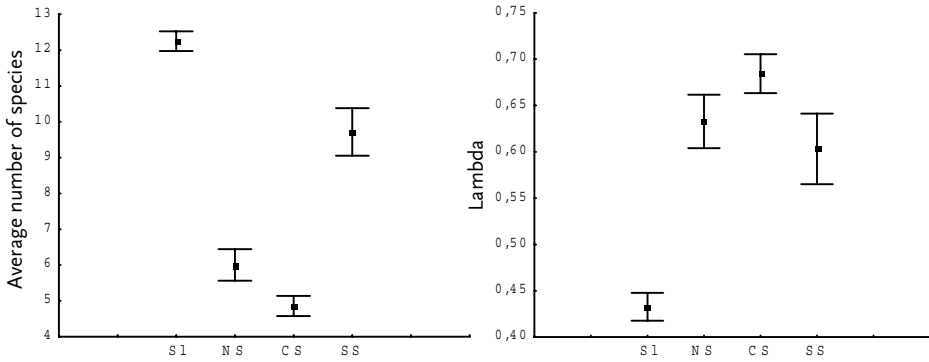


Figure 3.  $\alpha$  – Diversity: Average species richness (left) and dominance (right). S1, Slope; NS, Northern Shelf; CS, Central Shelf; SS, Southern Shelf. Mean  $\pm$ 0.95 Confidence Interval.

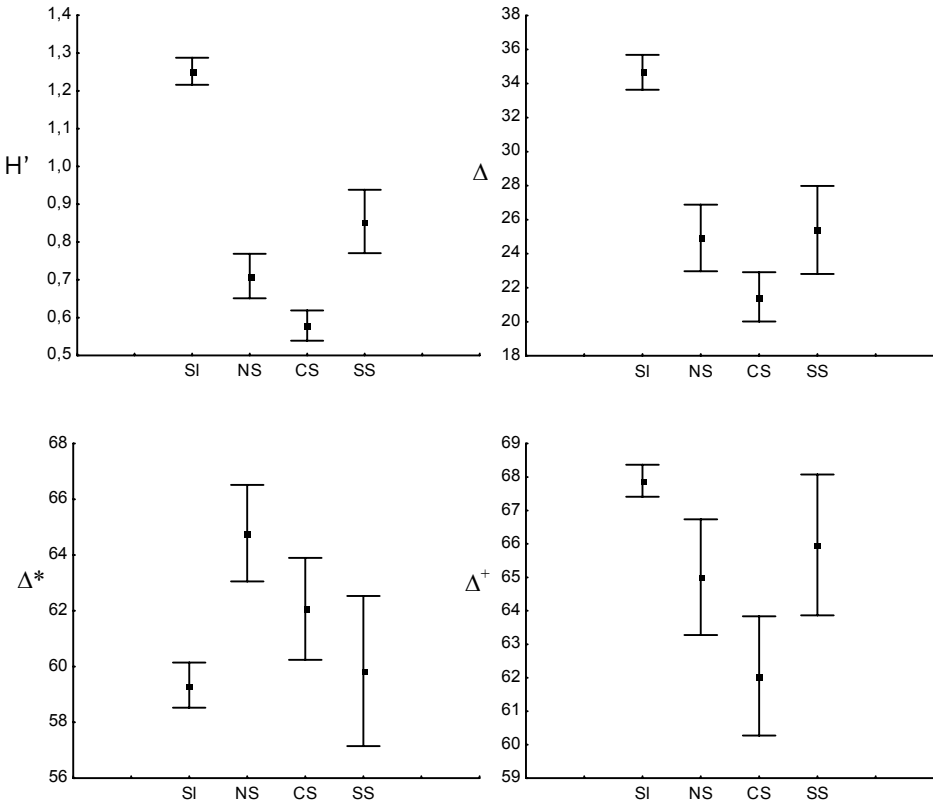


Figure 4.  $\alpha$  – Diversity. Shannon–Wiener,  $H'$ ,(upper left); average taxonomic diversity,  $\Delta$  (upper right); average taxonomic distinctness,  $\Delta^*$  (lower left); and average taxonomic breadth,  $\Delta^+$  (lower right). S1, Slope; NS, Northern Shelf; CS, Central Shelf; SS, Southern Shelf. Mean  $\pm$ 0.95 Confidence Interval.

indices ( $H'$  and  $\Delta$ ) show a pattern of higher diversity in the upper slope area as compared with the shelf communities, with the central slope having the lowest diversity values. This result is consistent with the patterns in richness and dominance described above. As regards taxonomic indices where the dominance component has been removed ( $\Delta^*$  and  $\Delta^+$ ), differences are not as marked and trends different. Remarkably, taxonomic distinctness is significantly higher in the northern and central shelves as compared with the slope community while taxonomic breadth is still highest in the slope area. The step lengths applied for the taxonomic indices are shown in Table 5.

### $\gamma$ -diversity

Figure 5 shows the species-area plots by major assemblage while Figure 6 presents the species-area plot for the three shelf communities combined.

Number of stations varies from area to area but clearly species richness is higher on the upper slope than in any of the other regions. Despite the very high number of samples taken here ( $> 1000$ ) the cumulative species curve does not seem to reach the asymptote, and therefore the total number of species may be expected to increase as sampling goes on. Comparison between shelf and upper slope (Figures 5, upper left and 6) shows how species richness is higher on the upper slope also when the three shelf communities are combined and number of samples is comparable between the two regions.

Table 5. Levels of classification used ( $k$ ) and weights (step lengths) applied using the 'standard' weighting scheme ( $\omega_k$ ) and an alternative scheme ( $\omega_k^0$ ) that uses an accumulation of the proportional decrease in taxon richness values ( $s_k$ ).

$k$	Level	$s_k$	$\omega_k$	$\omega_k^0$
1	Species	308	7.14	6.02
2	Genus	232	14.29	18.88
3	Family	112	21.43	34.87
4	Suborder	40	28.57	38.03
5	Order	35	35.71	52.52
6	Series	15	42.86	56.04
7	Superorder	13	50	70.42
8	Infradivision	6	57.14	75.35
9	Subdivision	5	64.29	75.35
10	Division	5	71.43	75.35
11	Subclass	5	78.57	75.35
12	Class	5	85.71	87.68
13	Subphylum	2	92.86	100
14	Phylum	1	100	100

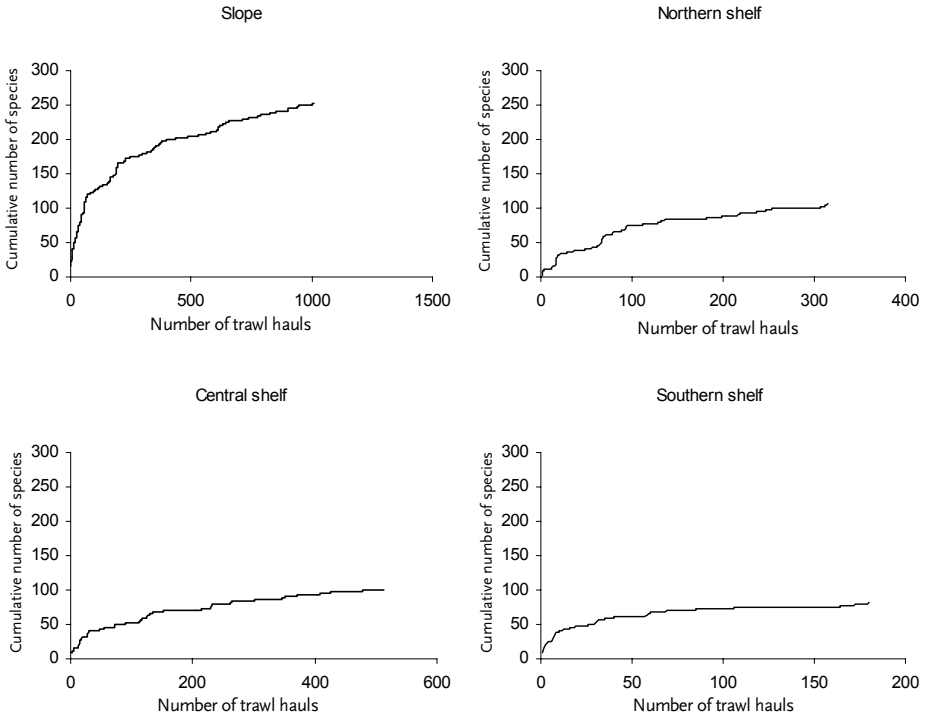


Figure 5. Species-area plot including all stations sampled during the study period, by main habitat type.

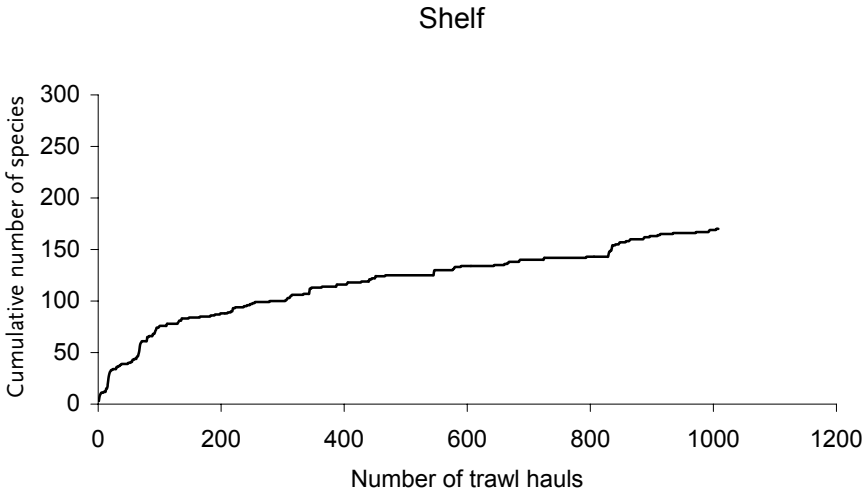


Figure 6. Species-area plot for the three shelf communities combined.

Table 6 shows a summary with the  $\gamma$ -diversity indices calculated for each of the assemblages and including all the stations sampled during the study period. These values provide therefore an average total diversity for the communities over a 10 year period.

$\gamma$ -diversity calculated by the Shannon-Wiener index gives the highest value for the slope area and the lowest for the central shelf area. This correlates with a high dominance ( $\lambda$ ) on the central shelf and a high evenness on the upper slope and is consistent with the results obtained at the  $\alpha$ -diversity level. The same pattern is shown in  $\Delta$ , but for  $\Delta^*$  the central shelf has the highest value and the three other areas are not significantly different from each other.

Figure 7 shows the 95% confidence intervals for the four regions of the mean taxonomic breadth. The mean is the theoretical mean from the overall value of  $\Delta^+$  for the global species list from the whole region while the variances are obtained by sampling a varying number of species (without replacement) from the total list of species. This produces an overall 'confidence funnel' that allows to check to what extent taxonomic breadth of a given set of samples falls within the expected values. The results show that most of the samples fall within the expected range but that most of them were below the expected mean.

## DISCUSSION

### Data limitations

Fish diversity studies have important limitations due to the difficulties of sampling a three-dimensional environment. Trawl gear, although among the least selective gears, can only sample a small fraction of the water column at the time, is size selective (but possibly this problem is less important on the RV Dr. Fridtjof Nansen, given that double lining in the codend and very small mesh sizes are used) and is species selective (some species living very close to the bottom might not be sampled properly). It is difficult to evaluate how this affects the various measures of diversity. It may be particularly im-

Table 6.  $\gamma$ -diversity with taxonomic diversity indices weighed for species richness.

Area	S	H'(loge)	Lambda	$\Delta$	$\Delta^*$	$\Delta^+$
Slope	252	3.12	0.07	57.11	61.25	72.77
Northern shelf	106	2.01	0.17	50.91	61.65	71.21
Central shelf	99	1.19	0.40	41.83	69.45	72.34
Southern shelf	97	2.13	0.18	50.36	61.75	72.40

portant for the measures that have an equity component given that the relative abundance of species found in the catches does not necessarily reflect the actual relative abundance. These inaccuracies are further exacerbated by fish behaviour. Although the various species are described in the broad categories “demersal” or “pelagic”, most carry out vertical diel migrations, more or less extensive depending on the species and/or the environmental conditions. Most of the species included in the analyses show this behaviour, but species that are mainly pelagic have been excluded from the analyses. For this reason, the study does not provide a comprehensive description of the total fish fauna found in the area.

### Patterns of fish species diversity on the Namibian shelf and upper slope areas

The Namibian shelf and upper slope belong to the temperate South African region. Its northern limit coincides with the Angola~Benguela frontal area, situated off southern Angola where the tropical Eastern Atlantic zoogeographic region has its southern limit. This region is the poorest in fish species of all tropical regions, with 434 shore fishes (as compared, for example, with 900 on the other side of the Atlantic). The South African region, which includes both the Indian Ocean and Atlantic sides of southern Africa, has a higher number of species given the presence of warm currents that

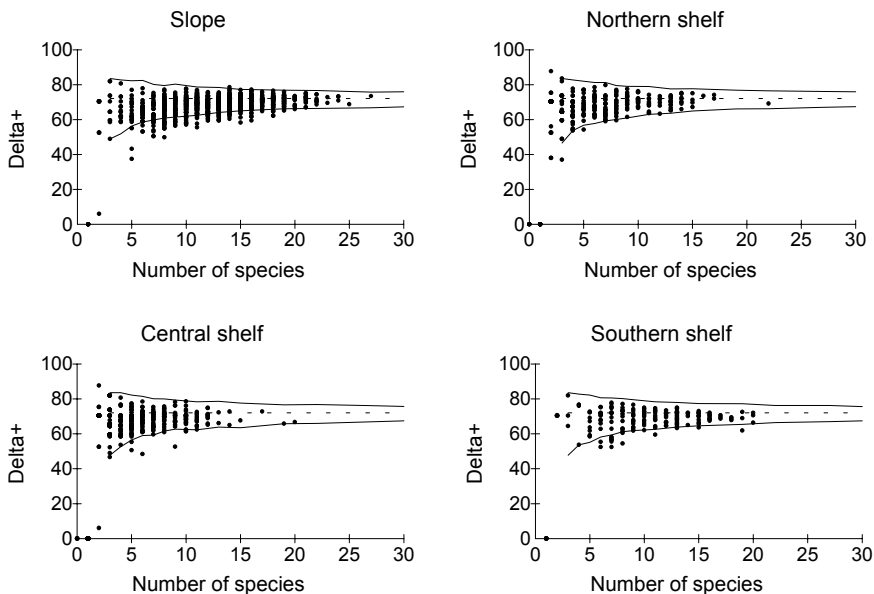


Figure 7. Funnel plots showing the average taxonomic distinctness of samples within each community, as compared with the 95% probability limits for  $\Delta^+$  and the mean taxonomic distinctness for the whole area.

extend the tropical Indo-Pacific fauna off the eastern and southern coasts of South Africa. Given its long isolation, this region has a high level of endemism. Namibia is part of the Southwestern Africa province (Briggs, 1974) that extends from southern Angola to the Cape of Good Hope.

Shelf diversity off Namibia is relatively low, with a total of 170 species recorded from 1008 trawl hauls. Only an average of 5 species per trawl haul is caught in the central shelf where near-bottom oxygen levels are lowest. Species richness at  $\alpha$ -diversity level on the shelf reflects the poor ecological conditions in the central shelf, while the trends in the northern and southern shelves reflect the diversity of adjacent areas. Species richness measured as average number of species per trawl haul is highest on the slope. Both of the above trends seem to be in contrast with two accepted rules in marine zoogeography: 1) that fish richness increases with decreasing latitude and; 2) that richness decreases with depth. Macpherson and Duarte (1994) have shown for the Eastern Atlantic that species richness declines by about 1% for each degree of latitude. Obviously this seems not to apply locally for the Namibian marine fishes. Richness and dominance are negatively correlated (Figure 3), with high and low richness corresponding to low and high dominance, respectively. High dominance and low diversity are typical of 'stressful' environments and the low oxygen levels found in large parts of the shelf area off Namibia are likely the reason for the observed patterns. Other studies have shown how fish diversity is strongly affected by oxygen levels  $< 1 \text{ ml l}^{-1}$ . (Bianchi 1991 and 1992).

Taxonomic indices showed similar trends to the species indices. In particular, the average taxonomic diversity ( $\Delta$ ) and the average taxonomic breadth ( $\Delta^+$ ) showed similar patterns to the Shannon-Wiener index, thus apparently not adding more information and insight, similarly to what was concluded by Hall and Greenstreet (1998) for patterns of change in the fish communities of the northern North Sea. However, taxonomic indices present a new aspect of diversity that seems worthwhile pursuing. Despite the higher species richness of the slope, its taxonomic range is similar to the shelf regions. Furthermore, diversity of higher taxa (expressed as ratio of number of higher taxa divided by the number of species in the assemblage) is in fact higher on the shelf than in the upper slope area (Figure 2). Two conclusions can be derived from this. The first is that relatively higher taxonomic diversity on the shelf may be related to higher ecological diversity. On the other hand, high taxonomic diversity associated with low species diversity at taxon level, may also result in ecologically unstable conditions. For example, the central shelf area holds relatively few species and has high levels of dominance. This may indicate the presence of (keystone?) fish species particularly well adapted to the given conditions but whose function in the ecosystem may be difficult to replace by other species in the case of popula-

tion crashes due, for example, to overexploitation. In these cases of low species diversity in stressful environments, resilience of the system may be low and the precautionary approach to fisheries management should be more strongly emphasized.

Of the three taxonomic indices used in this study ( $\Delta$ ,  $\Delta^*$  and  $\Delta^+$ ),  $\Delta^*$  has been difficult to interpret. It displays a quite different pattern as compared with all the other indices, but is consistent with the analysis of diversity of higher taxa shown in Figure 2.

Visualization of trends of taxonomic breadth within each community as compared with the theoretical mean is indicative of a lower local taxonomic breadth at community level as compared with the total. This confirms the distinctness of the four regions. It is suggested that diversity studies in a given geographic area should be preceded by analyses that test the homogeneity of the area, by, for example, multivariate analyses. Correlation with environmental variables leads to the definition of habitats, thus allowing separation of environmental effects from anthropogenic effects in understanding patterns of diversity.

### **Demersal fish diversity and fisheries management - A pragmatic approach toward monitoring fish diversity**

Clearly diversity has many facets and can hardly be represented by a single index. The question still remains as to which indices should be used to indicate ecosystem health as related to possible effects of fishing, and what are the reference points for these measures that fisheries management should aim at. Based on the experience made through the work of the SCOR working group on ecosystem effects of fishing (ICES, 2000) and the present study, the following is an outline for a pragmatic approach to monitoring demersal marine fish diversity.

Bottom trawl surveys, usually carried out under controlled conditions and with appropriate sampling schemes, are performed in areas exploited by the industrial fisheries. This type of data could be used to study diversity patterns in space and time, according to the following analytical steps:

- Stratification of a given area into communities by multivariate analyses and habitat definition (by, for example, main environmental variables). This would provide a definition of ecological and management units.
- Analysis of community properties as regards diversity, taking into account species richness, dominance and measures of taxonomic diversity.
- Monitoring trends over time. Development of reference points should be done in an adaptive way, at least to start with.

Methods should be standardized so that experiences gained in a given area could more easily be compared. Standardization of methods is a key issue and should be sought for all the analytical steps. For example, when looking



at taxonomic indices, it is essential that the number of taxonomic levels and the type of taxa utilized are the same given that the choice of number and type of taxa affects the taxonomic index. Furthermore, there are other features of diversity that may be important to evaluate (e.g. functional diversity) and studies should be carried out to find which of the indices are most informative in relation to assessing ecosystem health.

It is recommended that international (e.g. FAO) and regional fisheries organizations should take the lead as regards promoting the standardization of methods for describing and monitoring marine fish diversity.

## REFERENCES

- Bianchi, G. (1991): Demersal assemblages of the continental shelf and upper slope edge between the Gulf of Tehuantepec (Mexico) and the Gulf of Papagayo (Costa Rica). *Marine Ecology Progress Series* 73:121-140.
- Bianchi, G. (1992): Demersal assemblages of the continental shelf and upper slope of Angola. *Marine Ecology Progress Series* 81:101-120.
- Bianchi, G., Carpenter, K.E., Roux, J-P., Molloy, F.J., Boyer, D. and Boyer, H. (1993): FAO species identification field guide for fisheries purposes. The living marine resources of Namibia. Rome, FAO. 250 pp., 8 colour plates.
- Bianchi, G., Carpenter, K.E., Roux, J-P., Molloy, F.J., Boyer, D. and Boyer, H. (1999): FAO species identification field guide for fisheries purposes. *The living marine resources of Namibia*. 2<sup>nd</sup> edition. Rome, FAO. 265 pp., 11 colour plates.
- Bianchi, G., Hamukuaya, H. and Alveheim, O. (2001): On the dynamics of demersal fish assemblages off Namibia in the 1990s. In: A.I.L. Payne, S.C. Pillar, and R.J.M. Crawford (eds.). *South African Journal of Marine Science* 23: 419-428.
- Briggs, J.C. (1974): *Marine Zoogeography*. McGraw-Hill Book Company. 475pp.
- Clarke, K.R. and Warwick, R.M. (1998): A taxonomic distinctness index and its statistical properties. *Journal of Applied Ecology* 35:523-531.
- Clarke, K.R. and Warwick, R.M. (1999): The taxonomic distinctness measure of biodiversity: weighing of step lengths between hierarchical levels. *Marine Ecology Progress Series*, 184: 21-29.
- Clarke, K.R. and Warwick, R.M. (2001a): A further biodiversity index applicable to species lists: variation in taxonomic distinctness. *Marine Ecology Progress Series* 216: 265-278.
- Clarke, K.R. and Warwick, R.M. (2001b): *Change in marine communities: an approach to statistical analysis and interpretation*, 2<sup>nd</sup> edition. PRIMER-E: Plymouth.
- Fischer, W.G., Bianchi, G. and Scott, W.B. (eds.) (1981): FAO species identification sheets for fisheries purposes. Eastern Central Atlantic; fishing areas 34, 47 (in part). Rome, Food and Agricultural Organisation of the United Nations, vols 1-7: pag. var.
- Froese, R. and Pauly, D. (eds.) (2003): FishBase. World Wide Web electronic publication: [www.fishbase.org](http://www.fishbase.org), version 26 February 2003.

- Hall, S.J. and Greenstreet, S. P. (1998): Taxonomic distinctness and diversity measures: responses in marine fish communities. *Marine Ecology Progress Series* 166:227-229.
- Hamukuaya, H., Bianchi, G. and Baird, D. (2000): The structure of demersal assemblages off Namibia in relation to abiotic factors. *South African Journal of Marine Science* 23:397-417.
- ICES (2000): Ecosystem Effects of Fishing. Proceedings of an ICES/SCOR Symposium held in Montpellier, France 16-19 March 1999. ICES Marine Science Symposia Vol. 57, no. 3.
- Jennings, S. and Reynolds, J.D. (2000): Impacts of fishing on diversity: from pattern to process. In: *The Effects of Fishing on Non-target Species and Habitats* (M.J. Kaiser and S.J. de Groot, eds.), pp. 235-250.
- Macpherson, E. and Duarte, C.M. (1994): Patterns in species richness, size, and latitudinal range of East Atlantic fishes. *Ecography* 17:242-248.
- Moyle, P.B. and Cech, J.J. (1982): *Fishes. An Introduction to Ichthyology*. Prentice-Hall International (UK) Limited, London. 612 pp.
- Rogers, S.I., Clarke, K.R. and Reynolds, J.D. (1999): The taxonomic distinctness of coastal bottom-dwelling fish communities of the North-east Atlantic. *Journal of Animal Ecology* 68: 769-782.
- Smith, M. M. and Heemstra P.C. (eds.) (1991): *Smiths' Sea Fishes*. Southern Book Publishers, Johannesburg. 1048 pp.
- Strømme, T. (1992): NAN-SIS: Software for fishery survey data logging and analysis. User's manual. FAO. Computerised information services (Fisheries) 4. Rome. 103 pp.
- Warwick, R.M. and Clarke, K.R. (1995): New 'biodiversity' measures reveal a decrease in taxonomic distinctness with increasing stress. *Marine Ecology Progress Series* 129, 301-305.
- Warwick, R.M. and Clarke, K.R. (2001): Practical measures of marine biodiversity based on relatedness of species. *Oceanography and Marine Biology: an Annual Review* 39: 207-231.
- Whittaker, R.H. (1960): Vegetation of the Siskiyou Mountains, Oregon and California. *Ecological Monographs* 30: 279-338.

**APPENDIX 1. List of fish families and species collected by bottom trawl by the RV DR. F. Nansen (1990-2000).**

Family	Species	Family	Species
HAGFISHES			<i>Scymnodon squamulosus</i>
Myxinidae	<i>Myxine capensis</i>		<i>Isistius brasiliensis</i>
			<i>Oxynotus centrina</i>
		Centrophoridae	
CARTILAGINEOUS FISHES			<i>Centrophorus</i> sp.
Lamnidae	<i>Isurus oxyrinchus</i>		<i>Centrophorus uyato</i>
	<i>Apristurus</i> sp.		<i>Centrophorus granulatus</i>
	<i>Apristurus saldanha</i>		<i>Centrophorus squamosus</i>
	<i>Galeus polli</i>		<i>Deania profundorum</i>
Scyliorhinidae	<i>Holohalaelurus regani</i>		<i>Deania calcea</i>
	<i>Scyliorhinus capensis</i>		<i>Deania quadrispinosum</i>
	<i>Galeorhinus galeus</i>	Squalidae	<i>Squalus blainvillei</i>
Triakidae	<i>Mustelus palumbes</i>		<i>Squalus megalops</i>
	<i>Mustelus mustelus</i>		<i>Squalus acanthias</i>
			<i>Squalus mitsukurii</i>
Carcharhinidae	<i>Carcharhinus signatus</i>	Rajidae	<i>Raja</i> sp.
	<i>Prionace glauca</i>		<i>Raja miraletus</i>
Chlamydoselachidae	<i>Chlamydoselachus</i> <i>anguineus</i>		<i>Raja caudaspinosa</i>
	<i>Heptranchias perlo</i>		<i>Raja alba</i>
Hexanchidae	<i>Hexanchus griseus</i>		<i>Raja straeleni</i>
			<i>Raja wallacei</i>
Echinorhinidae	<i>Echinorhinus brucus</i>		<i>Raja clavata</i>
			<i>Raja stenorhynchus</i>
Dalatiidae	<i>Etmopterus</i> sp.		<i>Raja leopardus</i>
	<i>Etmopterus pusillus</i>		<i>Raja confundens</i>
	<i>Etmopterus lucifer</i>		<i>Raja doutrei</i>
	<i>Etmopterus spinax</i>		<i>Raja pullopunctata</i>
	<i>Etmopterus brachyurus</i>		<i>Raja springeri</i>
	<i>Centroscymnus crepidater</i>		<i>Raja spinacidermis</i>
	<i>Centroscymnus coelolepis</i>		<i>Cruriraja parcomaculata</i>
	<i>Centroscyllium fabricii</i>		<i>Bathyraja smithii</i>
		Torpedinidae	<i>Torpedo nobiliana</i>
		Dasyatidae	<i>Dasyatis</i> sp.

Family	Species	Family	Species
Myliobatidae			<i>Xenodermichthys copei</i>
	<i>Myliobatis aquila</i>	Platyroctidae	
Callorhynchidae			<i>Maulisia microlepis</i>
	<i>Callorhynchus capensis</i>	Gonostomatidae	
Rhinochimaeridae			<i>Diplophos</i> sp.
	<i>Neoharriotta pinnata</i>		<i>Diplophos maderensis</i>
Chimaeridae			<i>Triplophos</i> sp.
	<i>Hydrolagus</i> sp.		<i>Triplophos hemingi</i>
	<i>Hydrolagus mirabilis</i>	Sternoptychidae	
BONY FISHES			<i>Argyropelecus aculeatus</i>
Albulidae			<i>Argyropelecus affinis</i>
	<i>Pterothrissus belloci</i>		<i>Maurolucus muelleri</i>
Colocongridae		Phosichthyidae	
	<i>Coloconger cadenati</i>		<i>Phosichthys</i> sp.
Muraenidae			<i>Phosichthys argenteus</i>
	<i>Uropterygius wheeleri</i>		<i>Yarrella blackfordi</i>
Ophichthidae		Stomiidae	
	<i>Mystriophis rostellatus</i>		<i>Melanostomias</i> sp.
	<i>Ophisurus serpens</i>		<i>Odontostomias micropogon</i>
Nemichthyidae			<i>Photonectes braueri</i>
	<i>Nemichthys scolopaceus</i>		<i>Stomias boa boa</i>
	<i>Nemichthys curvirostris</i>		<i>Astronesthes</i> sp.
Congridae			<i>Bassanago albescens</i>
	<i>Bathyroconger vicinus</i>	Ateleopodidae	
Nettastomatidae			<i>Guentherus altivela</i>
	<i>Venefica proboscidea</i>	Chlorophthalmidae	
Synaphobranchidae			<i>Chlorophthalmus</i>
	<i>Synaphobranchus kaupii</i>		<i>atlanticus</i>
Halosauridae			<i>Chlorophthalmus agassizi</i>
	<i>Halosaurus ovenii</i>		<i>Chlorophthalmus</i>
Notacanthidae			<i>punctatus</i>
	<i>Notacanthus sexspinis</i>	Notosudidae	
Microstomatidae			<i>Scopelosaurus meadi</i>
	<i>Nansenia</i> sp.	Paralepididae	
Bathylagidae			<i>Lestidium atlanticum</i>
	<i>Bathylagus glaucilis</i>		<i>Lestrolepis intermedia</i>
Alepocephalidae			<i>Lestidiops</i> sp.
	<i>Alepocephalus</i> sp.		<i>Macroparalepis</i>
	<i>Alepocephalus rostratus</i>		<i>macrogeneion</i>
			<i>Macroparalepis affinis</i>

Family	Species	Family	Species
Neoscolopelidae	<i>Neoscolopelus macrolepidotus</i>		<i>Physiculus capensis</i> <i>Tripteroptychys gilchristi</i>
Myctophidae	<i>Diaphus</i> sp. <i>Diaphus dumerili</i> <i>Diaphus hudsoni</i> <i>Lampadena</i> sp. <i>Lampadena pontifex</i> <i>Lampanyctodes hectoris</i> <i>Lampanyctus australis</i> <i>Symbolophorus boops</i>	Bregmacerotidae	<i>Bregmaceros</i> sp. <i>Merluccius</i> sp. <i>Merluccius polli</i> <i>Merluccius capensis</i>
Ophidiidae	<i>Ophidion</i> sp. <i>Dicrolene intronigra</i> <i>Brotula barbata</i> <i>Genypterus capensis</i> <i>Lamprogrammus exutus</i> <i>Selachophidium guentheri</i>	Merlucciidae	<i>Merluccius paradoxus</i> <i>Lyconus pinnatus</i>
Macrouridae	<i>Coelorinchus</i> sp. <i>Coelorinchus fasciatus</i> <i>Coelorinchus coelorhinc.</i> <i>poll</i> <i>Coelorinchus braueri</i> <i>Coelorinchus matamua</i> <i>Coelorinchus acanthiger</i> <i>Hymenocephalus italicus</i> <i>Malacocephalus laevis</i> <i>Malacocephalus</i> <i>occidentalis</i> <i>Nezumia</i> sp. <i>Nezumia aequalis</i> <i>Nezumia leonis</i> <i>Nezumia milleri</i> <i>Nezumia micronychodon</i> <i>Trachyrincus scabrurus</i> <i>Gadella imberbis</i> <i>Laemonema laureysi</i>	Batrachoididae	<i>Batrachoides</i> sp. <i>Chatrabus melanurus</i> <i>Perulibatrachus rossignoli</i>
		Lophiidae	<i>Lophius piscatorius</i> <i>Lophius vaillanti</i> <i>Lophius vomerinus</i>
		Chaunacidae	<i>Chaunax pictus</i>
		Ogcocephalidae	<i>Dibranchus atlanticus</i>
		Melanocetidae	<i>Melanocetus johnsoni</i>
		Diceratiidae	<i>Phrynichthys wedli</i>
		Ceratiidae	<i>Ceratias holboelli</i> <i>Cryptosaras couesii</i>
		Scomberesocidae	<i>Scomberesox saurus</i>
		Trachipteridae	<i>Trachipterus trachipterus</i> <i>Trachipterus jacksonensis</i> <i>Zu elongatus</i>
		Regalecidae	<i>Regalecus glesne</i>
Moridae	<i>Lepidion capensis</i>	Trachichthyidae	<i>Hoplostethus cadenati</i> <i>Hoplostethus mediterraneus</i>

Family	Species	Family	Species
	<i>Hoplostethus atlanticus</i>	Acropomatidae	
	<i>Hoplostethus melanopus</i>		<i>Synagrops microlepis</i>
Berycidae	<i>Beryx splendens</i>	Polyprionidae	<i>Polyprion americanus</i>
Barbourisiidae	<i>Barbourisia rufa</i>	Callanthiidae	<i>Callanthias legras</i>
	<i>Cyttus traversi</i>	Serranidae	<i>Anthias anthias</i>
Zeidae	<i>Zeus faber</i>	Epigonidae	<i>Epigonus</i> sp.
	<i>Zeus capensis</i>		<i>Epigonus telescopus</i>
	<i>Zenopsis conchifer</i>		<i>Epigonus pandionis</i>
Oreosomatidae			<i>Epigonus denticulatus</i>
	<i>Allocyttus verrucosus</i>	Bramidae	
	<i>Neocyttus rhomboidalis</i>		<i>Brama brama</i>
	<i>Oreosoma atlanticum</i>		<i>Taractichthys longipinnis</i>
Grammicolepididae			<i>Taractes</i> sp.
	<i>Xenolepidichthys dagleishi</i>	Caristiidae	<i>Caristius groenlandicus</i>
Centriscidae	<i>Macrorhamphosus scolopax</i>	Emmelichthyidae	<i>Emmelichthys nitidus</i>
	<i>Notopogon macrosolen</i>	Sparidae	<i>Dentex macrophthalmus</i>
Congiopodidae	<i>Congiopodus spinifer</i>		<i>Spondyliosoma cantharus</i>
	<i>Congiopodus torvus</i>		<i>Argyrosomus hololepidotus</i>
Liparidae	<i>Careproctus griseldea</i>	Sciaenidae	<i>Umbrina canariensis</i>
Scorpaenidae	<i>Scorpaena</i> sp.		<i>Atractoscion aequidens</i>
Psychrolutidae	<i>Ebinania costaecanarie</i>	Percophidae	<i>Bembrops heterurus</i>
	<i>Psychrolutes macrocephalus</i>	Callionymidae	<i>Paracallionymus costatus</i>
Setarchidae	<i>Ectreposebastes imus</i>	Gobiidae	<i>Sufflogobius bibarbatu</i>
Sebastidae	<i>Helicolenus dactylopterus</i>	Sphyraenidae	<i>Sphyraena guachancho</i>
	<i>Sebastes capensis</i>	Gempylidae	<i>Paradiplospinus gracilis</i>
	<i>Trachyscorpia capensis</i>		<i>Ruvettus pretiosus</i>
Triglidae	<i>Chelidonichthys capensis</i>		<i>Thyrsites atun</i>
	<i>Chelidonichthys queketti</i>		
	<i>Trigla lyra</i>		

<b>Family</b>	<b>Species</b>	<b>Family</b>	<b>Species</b>
Trichiuridae	<i>Aphanopus</i> sp. <i>Benthodesmus tenuis</i> <i>Lepidopus caudatus</i> <i>Trichiurus</i> sp. <i>Trichiurus lepturus</i> <i>Scomber japonicus</i>	Tetragonuridae	<i>Tetragonurus cuvieri</i> <i>Tetragonurus atlanticus</i>
Scombridae	<i>Allothunnus fallai</i>	Bothidae	<i>Arnoglossus imperialis</i> <i>Arnoglossus capensis</i> <i>Monolene microstoma</i>
Centrolophidae	<i>Centrolophus niger</i> <i>Hyperoglyphe moselii</i> <i>Schedophilus pamarco</i> <i>Schedophilus huttoni</i> <i>Schedophilus ovalis</i>	Soleidae	<i>Austroglossus microlepis</i> <i>Austroglossus pectoralis</i> <i>Dicologlossa cuneata</i> <i>Synaptura kleini</i>
Nomeidae	<i>Cubiceps caeruleus</i>	Cynoglossidae	<i>Cynoglossus capensis</i> <i>Cynoglossus zanzibarensis</i>
		Diodontidae	<i>Chilomycterus reticulatus</i>